



Enabling Legendary Discovery™

LEGEND MAX™

ELISA Kit with Pre-coated Plate



Human PlGF

Cat. No. 446007

ELISA Kit for Accurate Quantitation of Human PlGF from
Cell Culture Supernatant, Serum, and Urine

BioLegend, Inc.
biolegend.com

It is highly recommended that this manual be read in its entirety before using this product. Do not use this kit beyond the expiration date.

For Research Purposes Only. Not for use in diagnostic or therapeutic procedures. Purchase does not include or carry the right to resell or transfer this product either as a stand-alone product or as a component of another product. Any use of this product other than the permitted use without the express written authorization of BioLegend is strictly prohibited.

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LEGEND MAX™ Human PIGF ELISA Kit

Introduction:

Placental growth factor (PIGF) is a member of the PDGF/VEGF family of growth factors. Four alternatively-spliced isoforms have been identified: PIGF-1, PIGF-2, PIGF-3, and PIGF-4. All isoforms are expressed in placental tissues, with PIGF-2 being expressed specifically during early placental development. PIGF-1 and PIGF-3 are considered diffusible isoforms, while PIGF-2 and PIGF-4 each contain a unique heparin binding domain that may increase associations with cell membranes. In addition to placental tissues, PIGF is expressed in the microvasculature, bone marrow, umbilical vein endothelia, uterine keratinocytes, and NK cells. PIGF binds to both VEGF-R1 and neuropilin, but not VEGF-R2 and induces monocyte activation, migration, inflammatory cytokine production. PIGF plays a key role in maternal vascular function during pregnancy, with circulating levels reaching a peak mid-gestation. It can be used as a marker for pregnancy-associated hypertensive disorders such as pre-eclampsia. High levels of PIGF expression have also been linked to tumorigenesis; specifically PIGF-1 has been identified in both colon and mammary carcinomas.

The BioLegend LEGEND MAX™ Human PIGF ELISA Kit is a Sandwich Enzyme-Linked Immunosorbent Assay (ELISA) with a 96-well strip plate that is pre-coated with a monoclonal mouse anti-human PIGF capture antibody. The detection antibody is a biotinylated monoclonal mouse anti-human PIGF antibody. This kit is specifically designed for the accurate quantitation of human PIGF from serum, urine, cell culture supernatant and other biological fluids. It is analytically validated with ready-to-use reagents.

Materials Provided:

Description	Quantity	Volume (per bottle)	Part #
Anti-Human PIGF Pre-coated 96-well Strip Microplate	1 plate		75151
Human PIGF Detection Antibody	1 bottle	12 mL	75152
Human PIGF Standard	1 vial	lyophilized	75154
Matrix A (for serum samples only)	1 vial	lyophilized	78303
Avidin-HRP	1 bottle	12 mL	77897
Assay Buffer A	1 bottle	25 mL	78232
Wash Buffer (20X)	1 bottle	50 mL	78233
Substrate Solution F	1 bottle	12 mL	79132
Stop Solution	1 bottle	12 mL	79133
Plate Sealers	4 sheets		78101

Materials to be Provided by the End-User:

- Microplate reader able to measure absorbance at 450 nm
- Adjustable pipettes to measure volumes ranging from 1 µL to 1,000 µL
- Deionized water
- Wash bottle or automated microplate washer
- Log-Log graph paper or software for data analysis
- Polypropylene tubes to prepare standard dilutions
- Timer
- Plate Shaker
- Polypropylene vials

Storage Information:

Store unopened kit components between 2°C and 8°C. Do not use this kit beyond its expiration date.

Opened or Reconstituted Components	
Microplate wells	If not all microplate strips are used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal. Store between 2°C and 8°C for up to one month.
Standard	The remaining reconstituted standard stock solution can be aliquoted into polypropylene vials and stored at -70°C for up to one month. Avoid repeated freeze-thaw cycles.
Matrix A	
Detection Antibody	Store opened reagents between 2°C and 8°C and use within one month.
Avidin-HRP	
Assay Buffer A	
Wash Buffer (20X)	
Substrate Solution F	
Stop Solution	

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin. Refer to the MSDS online at BioLegend's website for details (www.biolegend.com/msds).
2. Substrate Solution F is harmful if inhaled or ingested. Avoid skin, eye and clothing contact.

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3. To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum, plasma and other biological fluids in accordance with NCCLS regulations.
4. Stop Solution contains strong acid. *Wear eye, hand, and face protection.*
5. Before disposing of the plate, rinse it with an excess amount of tap water.

Specimen Collection and Handling:

Specimens should be clear and non-hemolyzed. If possible, unknown samples should be run at a number of dilutions to determine the optimal dilution factor that will ensure accurate quantitation.

Cell Culture Supernatant: If necessary, centrifuge all samples to remove debris prior to analysis. It is recommended that samples be stored at < -70°C. Avoid repeated freeze-thaw cycles.

Serum: Use a serum separator tube and allow clotting for at least 30 minutes, then centrifuge for 10 minutes at 1,000 x *g*. Remove serum layer and assay immediately or store serum samples at < -70°C. Avoid repeated freeze-thaw cycles.

Urine: Collect the first urine of the day (mid-stream) directly into a sterile container. Centrifuge to remove particular matter. Assay immediately or aliquot and store samples at < -70°C. Avoid repeated freeze-thaw cycles.

Reagent and Sample Preparation:

Note: All reagents should be diluted immediately prior to use.

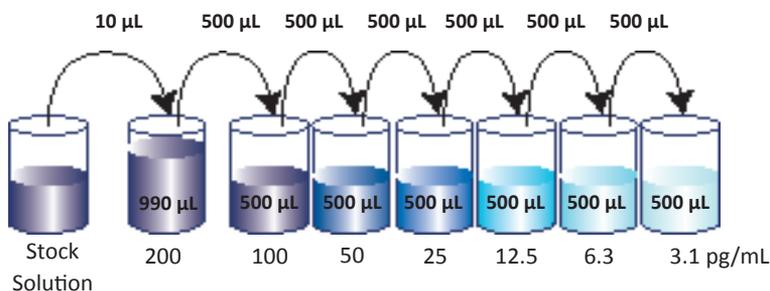
1. Dilute the 20X Wash Buffer to 1X with deionized water. For example, make 1 liter of 1X Wash Buffer by adding 50 mL of 20X Wash Buffer to 950 mL of deionized water. If crystals have formed in the 20X Wash Buffer, bring to room temperature and vortex until dissolved.
2. If serum samples will be assayed, reconstitute the lyophilized Matrix A by dispensing 2 mL of deionized water into the vial and allow the reconstituted Matrix A to sit at room temperature for 15 minutes, then vortex to mix completely.
3. Reconstitute the lyophilized Human PIGF Standard by adding the volume of Assay Buffer A to make the 20 ng/mL standard stock solution (Refer to LEGEND MAX Kit Lot-Specific Certificate of Analysis/LEGEND MAX Kit Protocol). Allow the reconstituted standard to sit at room temperature for 15 minutes, then briefly vortex to mix completely.

- In general, samples are analyzed without dilutions. However, if dilutions are required, use Assay Buffer A for diluting cell culture supernatant samples and urine samples and Matrix A for diluting serum samples.

Assay Procedure:

Note: Do not mix reagents from different kits or lots. Reagents and/or antibodies from different manufacturers should not be used with this kit.

- Bring all reagents to room temperature prior to use. It is strongly recommended that all standards and samples be run in duplicate or triplicate. A standard curve is required for each assay.
- If not all microplate strips will be used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal.
- Prepare 1,000 μL of the 200 pg/mL top standard by adding 10 μL of the standard stock solution in 990 μL of Assay Buffer A. Perform six two-fold serial dilutions of the 200 pg/mL top standard in separate tubes using Assay Buffer A as the diluent. Thus, the human PIGF standard concentrations in the tubes are 200 pg/mL , 100 pg/mL , 50 pg/mL , 25 pg/mL , 12.5 pg/mL , 6.3 pg/mL and 3.1 pg/mL , respectively. Assay Buffer A serves as the zero standard (0 pg/mL).



- Wash the plate 4 times with at least 300 μL of 1X Wash Buffer per well and blot any residual buffer by firmly tapping the plate upside down on absorbent paper. All subsequent washes should be performed similarly.
- For measuring urine samples or cell culture supernatant:**
 - Add 50 μL of Assay Buffer A to each well that will contain either standard dilutions or samples.
 - Add 50 μL of standard dilutions or samples to the appropriate wells.

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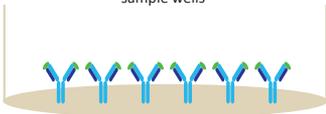
For measuring serum samples:

- a) Add 50 μL of Matrix A to each well that will contain the standard dilutions. Add 50 μL of Assay Buffer A to each well that will contain samples.
 - b) Add 50 μL of standard dilutions to the wells containing Matrix A. Add 50 μL of dserum samples to the wells containing Assay Buffer A.
6. Seal the plate with a Plate Sealer included in the kit. Shake the plate for 1 minute and incubate the plate **overnight** between 2°C and 8°C without shaking.
 7. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
 8. Add 100 μL of Human PIGF Detection Antibody solution to each well, seal the plate and incubate at room temperature for 2 hours while shaking.
 9. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
 10. Add 100 μL of Avidin-HRP solution to each well, seal the plate and incubate at room temperature for 30 minutes while shaking.
 11. Discard the contents of the plate into a sink, then wash the plate 5 times with 1X Wash Buffer as in step 4. For this final wash, soak wells in 1X Wash Buffer for 30 seconds to 1 minute for each wash. This will help minimize background.
 12. Add 100 μL of Substrate Solution F to each well and incubate for 15 minutes in the dark. Wells containing human PIGF should turn blue in color with an intensity proportional to its concentration. It is not necessary to seal the plate during this step.
 13. Stop the reaction by adding 100 μL of Stop Solution to each well. The solution color should change from blue to yellow.
 14. Read absorbance at 450 nm within 30 minutes. If the reader is capable of reading at 570 nm, the absorbance at 570 nm can be subtracted from the absorbance at 450 nm.

Assay Procedure Summary

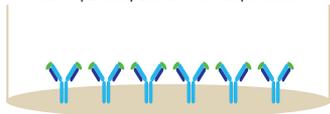
- For measuring urine or cell culture supernatant:**

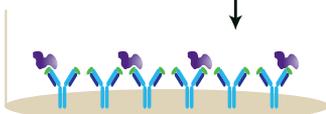
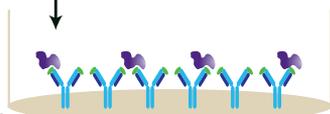
Wash 4 times
Add 50 μ L Assay Buffer A to standard and sample wells

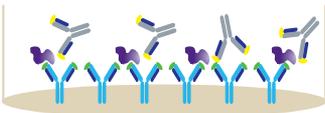


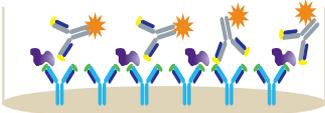
For measuring serum:

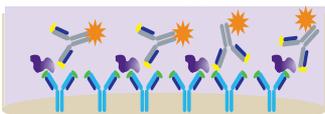
Wash 4 times
Add 50 μ L Matrix A to standard wells
Add 50 μ L Assay Buffer A to sample wells

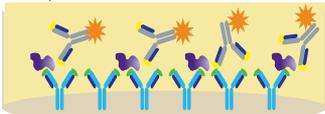

- Add 50 μ L diluted standards to standard wells
Add 50 μ L samples to sample wells
Shake for 1 min and incubate overnight between 2°C and 8°C without shaking



- Wash 4 times
Add 100 μ L Detection Antibody solution
Incubate 2 hr, RT, shaking


- Wash 4 times
Add 100 μ L Avidin-HRP A solution
Incubate 30 min, RT, shaking


- Wash 5 times
Add 100 μ L Substrate Solution F
Incubate 15 min, RT, in the dark


- Add 100 μ L Stop Solution


- Read absorbance at 450 nm and 570 nm

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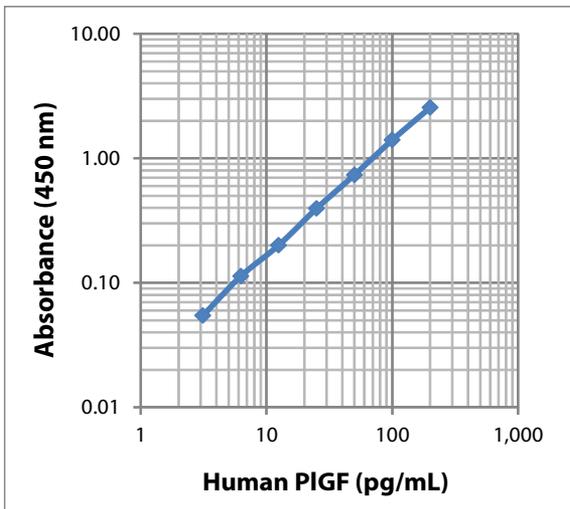
Calculation of Results:

The data can be best calculated with computer-based curve-fitting software using a 5- or 4-parameter logistics curve-fitting algorithm. If an appropriate software is not available, use log-log graph paper to determine sample concentrations. Determine the mean absorbance for each set of duplicate or triplicate standards, controls, and samples. Plot the standard curve on log-log graph paper with cytokine concentration on the X-axis and absorbance on the Y-axis. Draw a best fit line through the standard points. To determine the unknown cytokine concentrations, find the mean absorbance value of the unknown concentration on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the cytokine concentration.

If samples were diluted, multiply the concentration by the appropriate dilution factor. If a test sample's absorbance value falls outside the linear portion of the standard curve, the test sample needs to be re-analyzed at a higher (or lower) dilution as appropriate.

Typical Data:

This standard curve was generated at BioLegend for demonstration purposes only. A standard curve must be run with each assay.



Performance Characteristics:

Specificity: This kit recognizes natural and recombinant PIGF-1. No cross reactivity was observed when this kit was used to analyze the following recombinant cytokines/chemokines at 50 ng/mL.

Human	EGF, EPO, FASL, FGF-basic, G-CSF, GDNF, GM-CSF, IFN- α 2, IFN- γ , IGF-I, IGF-II, IL-1 α , IL-1 β , IL-1R, IL-2, IL-2Ra, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-11, IL-12(p70), IL-13, IL-15, IL-17A, IL-17A/F, IL-17F, IL-21, IL-22, IL-23, IL-27, IL-33, IL-34, LIF, LT- α , M-CSF, NGAL, PAXILLIN, PDGF-AA, PDGF-AB, PDGF-BB, PDGF-CC, PDGFRB Fc, SCF, TGF- α , TGF- β 1, TGF- β 2, TGF- β 3, TNF-RII, TNF- α , TSLP, VEGF-121, VEGF-165, VEGF-B167, VEGF-D, VEGFR1, VEGFR2-Fc chimera, β NGF
Mouse	CXC3CL1, EGF, EPO, GM-CSF, IFN- β 1, IFN- γ , IL-1 α , IL-1 β , IL-2, IL-3, IL-21, IL-22, IL-23, IL-25, IL-27, PDGF-BB, TNF- α , VEGF-120, VEGF-164

Recombinant PIGF-2 and PIGF-3 show 91.2% and 48.4% reactivity respectively with this assay. Recombinant human VEGF/PIGF heterodimer shows 0.12% cross-reactivity at 50 ng/mL. Recombinant human VEGFR1 starts to interfere at concentrations greater than 2000 pg/mL.

Sensitivity: The average minimum detectable concentration is 0.39 pg/mL.

Recovery: Recombinant human PIGF (100, 25 and 6.3 pg/mL) was spiked into human serum and urine samples and then analyzed with the LEGEND MAX™ Human PIGF ELISA kit.

Sample Type	N	% Recovery
Serum	5	88
Urine	4	85

Linearity: Recombinant human PIGF was spiked into human serum and urine samples to produce samples within the dynamic range and diluted up to 8-fold. Cell culture supernatant samples were diluted up to 320-fold. The samples were then assayed with the kit to determine the dilutional linearity.

Sample Type	N	% Linearity
Serum	5	100
Urine	4	108
Cell Culture Supernatant	2	100

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Intra-Assay Precision: Two serum samples were spiked with different recombinant human PIGF concentrations and tested with 16 replicates in one assay.

Concentration	Sample 1	Sample 2
Number of Replicates	16	16
Mean Concentration (pg/mL)	170.7	14.1
Standard Deviation	8.0	1.2
% CV	4.7	8.8

Inter-Assay Precision: Two serum samples were spiked with different concentrations of recombinant human PLGF and assayed on 5 different days.

Concentration	Sample 1	Sample 2
Number of Assays	5	5
Mean Concentration (pg/mL)	86.6	16.7
Standard Deviation	10.7	1.83
% CV	12.3	11.0

Normal Biological Samples:

Normal Ranges: Serum and urine samples from human donors were tested for endogenous human PIGF.

Sample Type	N	% Detectable	Min (pg/mL)	Max (pg/mL)	Mean (pg/mL)
Serum	26	100	4.1	30.1	13.1
Urine	9	100	6.3	40.8	23.1

Cell Supernatant: JAR (1×10^6 cells/mL) and JEG3 (1×10^6 cells/mL) cells were cultured. The cell supernatants were collected after 48 hours and assayed for human PIGF.

Cell Type	Concentration (pg/ml)
JAR	9173
JEG3	1367

Troubleshooting Guide:

Problem	Probable Cause	Solution
High Background	Background wells were contaminated	Avoid cross-well contamination by using the provided plate sealers. Use multichannel pipettes and change tips between pipetting samples and reagents.
	Insufficient washes	Increase number of washes. Increase soaking time between washes prior to addition of substrate solution.
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells. Use a clean container prior to pipetting substrate solution into wells.
No or poor signal	Detection Antibody, Avidin-HRP or Substrate solution were NOT added	Rerun the assay and follow the protocol.
	Wrong reagent or reagents were added in wrong sequential order	
	Insufficient plate agitation	The plate should be agitated during all incubation steps using a plate shaker at a speed where solutions in wells are within constant motion without splashing.
	The wash buffer contains Sodium Azide (NaN ₃)	Avoid Sodium Azide contamination in the wash buffer as it inhibits HRP activity.
	Incubations were done at an inappropriate temperature, timing or without agitation	Rerun the assay and follow the protocol.
Low or poor standard curve signal	The standard was incorrectly reconstituted or diluted	Adjust the calculations and follow the protocol.
	Standard was inappropriately stored	Store the reconstituted standard stock solution in polypropylene vials at -70°C. Avoid repeated freeze-thaw cycles.
	Reagents added to wells with incorrect concentrations	Check for pipetting errors and the correct reagent volume.

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Problem	Probable Cause	Solution
Signal is high, standard curves have saturated signal	Standard reconstituted with less volume than required	Reconstitute new lyophilized standard with the correct volume of solution recommended in the protocol.
	Standards/samples, detection antibody, Avidin-HRP or substrate solution were incubated for too long	Rerun the assay and follow the protocol.
Sample readings are out of range	Samples contain no or below detectable levels of the analyte	If samples are below detectable levels, it may be possible to use a larger sample volume. Contact technical support for appropriate protocol modifications.
	Samples contain analyte concentrations greater than highest standard point	Samples may require dilution and analysis.
High variation in samples and/or standards	Multichannel pipette errors	Confirm that pipette calibrations are accurate.
	Plate washing was not adequate or uniform	Ensure pipette tips are tightly secured. Ensure uniformity in all wash steps.
	Non-homogenous samples	Thoroughly mix samples before assaying.
	Samples may have high particulate matter	Remove particulate matter by centrifugation.
	Cross-well contamination	Do not reuse plate sealers. Always change tips for reagent additions. Ensure that pipette tips do not touch the reagents on the plate.

ELISA Plate Template

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												



The path to legendary discovery™

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